

## Are SNIP's still desirable in sports genomics?

MIROSLAV SLIŽIK<sup>1</sup>, BARBARA POSPIESZNA<sup>2</sup>, JOANNA GRONEK<sup>3</sup>, ROBERT SWOREK<sup>4</sup>

### Abstract

A single-nucleotide polymorphism (SNIP) is a variation in a single nucleotide that occurs at a certain position in the DNA. Each variant is, to some extent, present within a population (e.g. > 1%). Due to the correlations of some SNIP's with sport performance and athletic physical capacity, various authors considered their importance in the context of professional sport. Among many SNIP's angiotensin I converting enzyme (ACE) polymorphism is a well-studied example associated with an enhanced physiological response to aerobic exercise. Among other sport-related interesting SNIP's following are highly documented: AMPD1 (C34T) Gln12 Allele, BDKRB2 rs5810761, UCP's and eNOS rs1799983.

**KEYWORDS:** nucleotide polymorphism, sport performance, genetic predisposition.

Received: 16 December 2016

Accepted: 9 March 2017

Corresponding author: barpos@amu.edu.pl

<sup>1</sup> Matej Bel University in Banska Bystrica, Department of Physical Education and Sport, Banska Bystrica, Republic of Slovenia

<sup>2</sup> Adam Mickiewicz University in Poznań, Department of Tourism and Recreation, Poznań, Poland

<sup>3</sup> Poznań University of Physical Education, Department of Dance and Gymnastics, Poznań, Poland

<sup>4</sup> Poznań University of Physical Education, Division of Sports and Defence Education, Poznań, Poland

### Introduction

In highly professional sports – especially team sports such as soccer, American football, basketball, etc. – the market value of the players is skyrocketing, their value sometimes exceeds 100 million dollars. Such enormous financial funds spent by clubs on the athletes are forcing managers and club administration to collect far-reaching information on the player's psycho-physical profile. People who are responsible for such decisions are also very aware of the numerous potential risks associated with the high loads which are subjected to the athletes during both training and competition. In such environment there is also a big emphasis put on the role of genetic testing in the identification of predisposition to injury or other sudden episodes like Sudden Cardiac Death (SCD) while exercise. The detailed review covering the role of genetic testing in the identification of young athletes with inherited primitive cardiac disorders was prepared in 2016 by Tiziano et al. [1].

However, the question is what makes a champion – nurture or nature? Genes, environmental factors or gene-environment ( $G \times E$ ) interaction? Brutsaert and Parra have tried to answer this question [2]. The authors come up with the evidence to support the genetic basis to athletic performance, with some emphasis on the candidate gene studies. In their review they have definitely stressed environmental factors that influence the athletic performance and highlighting the irreversible environmental effects, i.e., epigenetic effects, fetal programming, or ones occurring during childhood and adolescence. The authors underline the significance of  $G \times E$  interaction in meaning of understanding variation

in human physiological performance [2]. Genes have a great impact on various athletic performance components such as strength, power, flexibility, neuromuscular coordination, endurance, psychological traits, and other phenotype traits. Athletes' condition is a heritable trait – it depends on sports discipline, but on average 66% of the variance in athletes' condition is explained by additive genetic factors. The residual variance is due to nonshared environmental factors.

The genetic studies related to sport origin from observations that identical twins engaging in competitive sports were significantly more likely to participate in the same sports than pairs of dizygotic twins [3]. The next documented step to identify genetic markers for sports performance relate to the Mexico (1968) and Montreal (1976) Olympic Games, yet the researchers did not generate any strong positive findings [4]. It was until 1998, when, the association between the ACE gene and an aptitude for sport was described by Montgomery et al. [5]. Since then our knowledge about the role of genetics in sports has changed significantly. Among others we have learned about SNIPs (Single Nucleotide Polymorphism's) and major genes (genes with major effect). Although the probability of becoming an elite athlete is very likely influenced by genetic factors only the few of the genes have been yet proven to be associated with motor skills [6, 7]. Nowadays there are many programs, which could be collectively called "Talent Search".

The scientists from the multiple-medal countries put the great expectations in the assessment of sport predisposition on the basis of Performance Enhancing Polymorphism (PEP's) [8]. However are genetic tests currently practical application as expectations are still significant? PEP's are those gene variants (the variants of genes) that may determine the critical for a given sport physiological features such as cell metabolism, muscular structure and even injury susceptibility. PEP's are based mainly on the SNIP's examination.

There are multiple single nucleotide polymorphisms in the human genes, raising the possibility that allelic differences in definite gene might influence physical performance [9], injury susceptibility [10] as well as nociception in the general population [11, 12]. SNIP's may affect levels of transcription, splicing, stability and expression of RNA by altering the amino acid sequence [13, 14, 15]. Are SNIP's still desirable in the study of genetic predisposition in sport? The answer to such question is positive, although the selection of adequate SNIP's remains a big challenge for researchers. In current research the most popular SNIP's are related

to: cardiorespiratory capabilities and skeletal muscle potential to exercise at high intensities, structure of muscle cells, aerobic and anaerobic power, injury susceptibility or sensitization to pain. Advanced research on those topics are carried out on a large scale in just a few countries in the world: the US, Russia, China, Australia, Spain, Israel, South Africa, Australia and Poland. Results published up to date have focused on differences in allele frequencies between athletes and non-athlete controls. Among thousands of major genes main interest was located on angiotensin I converting enzyme (*ACE*),  $\alpha$ -actinin-3 (*ACTN3*), and peroxisome proliferator-activated receptor- $\gamma$  coactivator 1 $\alpha$  (*PPARGC1A*) polymorphisms and on mitochondrial DNA (mtDNA) haplogroup distributions [16].

### **ACE (I/D)**

Angiotensin I converting enzyme (*ACE*) is a peptidase responsible for the blood pressure regulation, belonging to the renin-angiotensin system (RAS). *ACE* converts angiotensin I to angiotensin II, which is a very potent vasoconstricting factor [17]. Defective functioning of renin-angiotensin system may be the cause of numerous cardiovascular changes. Insertion-deletion (I/D) polymorphism within the *ACE* includes the two allelic variants characterized by presence/absence of the *Alu* repetitive 287 bp sequence in 16 intron. Thus *ACE* gene may have two alleles, distinctly different in their size: shorter – deletion allele (D) and longer – insertion allele (I). The activity of angiotensin I converting enzyme in the blood of individuals with DD genotype is about twice higher than in those with genotype II [18, 19], therefore the genotype II is correlated with a lower risk of cardiovascular disease [20]. Homozygous D/D, which often reveals elevated blood pressure, may be defined as a group under the risk of developing the cardiovascular system disease.

Insertion II genotype (homozygous insertion) has the low angiotensin activity in the tissues [18] and is associated with a better response to aerobic exercise [19]. It allows to maintain a favorable energy balance during the intense and prolonged physical exercise. It has been observed that athletes competing in disciplines with aerobic metabolism predominance, e.g. climbing, long-distance running, long distance swimming, almost never have the D allele in their genotype. In turn, athletes of anaerobic disciplines (with a predominance of anaerobic metabolism) – sprinters, short distance swimmers – are the ones with high levels of *ACE* and more frequent occurrence of DD genotype deletion [18, 19].

**AMPD1 (C34T) Gln12 Allele**

AMPD relocates the balance of the myokinase reaction in the ATP production process ( $2 \text{ ADP} \leftrightarrow \text{ATP} + \text{AMP}$ ) by transforming AMP to inosine monophosphate (IMP) [21, 22]. This reaction is important because of (i) rapid ATP synthesis (ii) AMP is potent stimulant for glycolysis [23]. AMPD is coded by three independent gene families (AMPD1 – is expressed in skeletal muscles, AMPD2 – is expressed in non-muscle tissue and smooth muscle, AMPD3 – is expressed in erythrocytes) [24]. The activity of AMP deaminase in myocytes is several times higher than its activity in other tissues – this condition is associated with the regulation of purine nucleotide cycle [25]. The AMPD reaction is the preliminary response of the purine nucleotide cycle and plays a central role in the recovery of adenine nucleotides [21]. AMPD, together with myokinase, participates in the ATP restoration in myocytes acting as muscle energy metabolism regulator during high-intensity exercise [26, 27, 28]. Physical exercises change muscle AMPD activity and *AMPD* expression in skeletal myocytes dependent on the fibre types [21, 22, 28]. Especially *AMPD1* is essentially expressed in fast-twitch muscle fibres where anaerobic activity causes a decrease in AMPD activity concurrent with an increase in the proportion of active fast-twitch (type II) fibres. Hence, AMPD expression appears to be influenced by the intensity of physical activity [29]. The nonsense mutation 34C>T (C to T transition in nucleotide 34, Gln12X, rs17602729) in exon 2 of the *AMPD1* gene converts glutamine codon (CAA) into the premature stop codon (TAA), and in consequences appears to be the main cause of AMPD deficiency [26, 27, 29]. Individuals with one normal and one mutant allele are more often engaged in intermediate activities, and those with two *AMPD1* normal alleles in high-intensity activities. The *AMPD1* CC genotype was found to be associated with anaerobic performance (Vertical Jump) [29]. Ginevičienė et al. [29] also found that the X allele is an unfavourable factor for athletes in sprint and power-oriented sports categories.

**BDKRB2**

The renin-angiotensin system (RAS) with its key component: angiotensin I converting enzyme (ACE), plays a fundamental role in circulation and blood homeostasis [30]. While ACE by vasoconstricting influence increases blood pressure, bradykinin (BK) by being a very potent endothelium-dependent vasodilator decreases it. In 1980, over 30 years after bradykinin was discovered by Maurício Rocha e Silva group, Regoli and Barabé proposed that BK acts via specific two cell-

surface receptors that are classified as the bradykinin 1 receptor (BDKRB1) and the bradykinin 2 receptor (BDKRB2) [31]. Both receptors are anchored on the plasma membrane of the myocytes and the vascular endothelium [32]. During physical activity BDKRB2 are consequently activated, what results in increased blood flow in the muscles, improved muscle glucose uptake, and thus higher endurance performance [33]. BDKRB2 is encoded by a single-copy gene, located to chromosome 14q32 and expressed in most human tissues. The insertion/deletion polymorphism (–9/+9, rs5810761) in exon 1 is the most commonly investigated polymorphism associated with athletes condition, as well as cardiovascular disease and hypertension [32, 34, 35]. Deletion of a 9 bp (–9) repeat in exon 1 of the *BDKRB2* gene is associated with higher mRNA expression, and increased receptor activity [36, 37]. It is suggested that –9 allele may be correlated with higher skeletal muscle metabolic adeptness and endurance performance [33]. The interesting research was conducted on swimmers with the –9/–9 genotype, who performed better in long distance competitions, than swimmers with other genotypes of the *BDKRB2* gene [30].

**UCP's**

ATP is produced by energy coupling, proceeding at the level of the electron transport chain in mitochondria. In adipose tissue, this coupling with ADP phosphorylation is only partial, because uncoupling proteins (UCPs) induce a proton leak, releasing the energy stored in ATP as heat [38]. Uncoupling proteins belong to the abundant family of mitochondrial anioncarrier proteins (MCAPs). Two of them may be taken into account as important for physical fitness: UCP2, which is expressed e.g. in muscles, lungs, spleen, heart, kidneys, central nervous system and white adipose tissue and UCP3, found in heart and skeletal muscles [39, 40]. The physiological role of UCP2 is not clear. Numerous studies showed that the most probable function of this protein is mild energy uncoupling, which accelerates metabolism and protects cells against damage by reducing the amount of reactive oxygen species (ROS) [38]. Fleury & Sanchis [39] and Bouchard et al. [41] supposed the UCP2 protein is associated with lipid metabolism and energy balance. Several SNIP's observed in *UCP2*, correlates with metabolic syndrome [42], obesity, BMI, resting metabolic rate [39, 43], or susceptibility to diabetes type 2 [44], but the results of those studies are equivocal. Some UCP2 SNIP's are associated with a higher energy efficiency, thus it is probable that UCP2 affects energy expenditure during physical activity [45, 46].

The decreased UCP3 expression (lower UCP3 mRNA level) was observed in athletes with negative correlation with  $VO_2\text{max}$ , therefore it is supposed that UCP3 expression may be influenced by strength training [39, 47]. Very regular physical activity or strength training decrease *UCP3* gene expression, and thus may increase energy efficiency in such athletes [39, 48].

Although lipid metabolism seems to be essential for aerobic capacity, so far only few studies have focused on the direct effects of polymorphisms of UCP genes on athletes performance [38, 49]. The authors analysed the association of the maximum oxygen uptake level ( $VO_2\text{max}$ ) with two polymorphisms: insertion/deletion (I/D) in exon 8 of the UCP2 gene and C>T substitution in exon 5 (630 C>T; Y210Y) of the UCP3 gene [38].

## NOS

Nitric oxide (NO), the molecule of the 1992 year, is produced by Nitric Oxide synthase (NOS; EC 1.14.13.39) from L-arginine – semiessential amino-acid derived from food, intracellular protein degradation or from endogenous synthesis [50, 51]. The L-arginine amine group is oxidized by molecular oxygen to L-citrulline and NO [52]. There are three isoforms of NOS: neuronal (nNOS or NOS I), inducible (iNOS or NOS II), and endothelial (eNOS or NOS III), which differ in the structure and function [50, 53]. Neuronal NOS is expressed in specific neurons of the central nervous system (CNS). It's action is associated with synaptic plasticity, central control of blood pressure and with penile erection [53]. Expression of the inducible NOS, which is  $Ca^{2+}$ -independent, can be stimulated in almost any cell or tissue, so long as there are appropriate inducing agents available – inflammatory mediators (e.g. cytokines) [52]. iNOS exhibit antibacterial effect due to generating the large amounts of NO which interacts with  $O_2^-$  leading to the local formation of toxic peroxynitrite ( $ONOO^-$ ). Beside the cytostatic effects excessive NO production by iNOS plays a crucial role in massive arteriolar vasodilatation seen in septic shock [54]. Endothelial NOS is expressed by endothelial cells, cardiac myocytes and cardiac conduction tissue [55]. Endothelial Nitric Oxide is a physiological vasodilator, but it also serve vasoprotective activity: it inhibits the platelet and leucocyte adhesion to the vascular wall and endothelial permeability avoiding the atherogenesis development or the release of platelet-derived growth factors preventing the smooth muscle proliferation. Moderate exercise leads to an improvement of endothelium function mainly through increment of the NOS activity, generally eNOS. Endothelial nitric oxide synthase (eNOS) is up-regulated by an increase in flow-mediated shear stress

associated with physical exercise, due to a complex pattern of intracellular regulations [56, 57, 58]. Investigations conducted on humans and animals have documented that exercise increases eNOS gene and protein expression [59, 60, 61]. Moreover under chronic exercise also the shear stress-induced eNOS phosphorylation occurs, so the ratio of phosphorylated to unphosphorylated eNOS gets higher in the trained individuals compared with the controls [58]. Therefore even without a significant increase in eNOS protein the improvement in functioning of the cardiovascular system may occur [62].

The gene is 21 kb of genomic DNA, 26 exons [63]. The most examined and functionally related common variants of the NOS3 are single nucleotide polymorphisms (SNP): 786T/C (rs2070744), G894T (Glu298Asp, rs1799983), as well as the intron 4 variable number tandem repeat (VNTR) [64]. Numerous studies indicate that –786T/C (rs2070744) and G894T NOS3 SNP's can be associated with several health/fitness, training or exercise response phenotypes e.g. adaptation of parasympathetic modulation response to exercise training, cardiovascular traits such as blood pressure, heart rate, cardio-biochemical parameters and vascular reactivity.

Other genes which still require to be investigated are: ACTN3, EPAS1, HIF1, IGF1, IL1 RN VNTR-86bp, IL-15, IL-6, MCT1, NFATC4, NRF1, PPAR $\alpha$ , PPAR $\gamma$ , PPP3R1, TFAM, TNF, VEGFA. In conclusion SNIPs are very informative and therefore seem to be useful in sports genomics. As shown in this paper some of them correlate with sport performance and athletes physical capacity.

## References

1. Tiziano FD, Palmieri V, Genuardi M, Zeppilli P. The role of genetic testing in the identification of young athletes with inherited primitive cardiac disorders at risk of exercise sudden death. *Front Cardiovasc Med.* 2016; 26(3): 28.
2. Brutsaert TD, Parra EJ. What makes a champion? Explaining variation in human athletic performance. *Respir Physiol Neurobiol.* 2006; 151(2-3): 109-123.
3. Gedda L. Sports and genetics. A study on twins (351 pairs). *Acta Genet Med Gemellol (Roma).* 1960; 9: 387-406.
4. Chagnon YC, Allard C, Bouchard C. Red blood cell genetic variation in Olympic endurance athletes. *J Sport Sci.* 1984; 2(2): 121-129.
5. Montgomery HE, Marshall R, Hemingway H, et al. Human gene for physical performance. *Nature.* 1998; 393: 221-222.
6. Pitsiladis Y, Wang G, Wolfarth B, et al. Genomics of elite sporting performance: what little we know and necessary advances. *Br J Sports Med.* 2013; 47(9): 550-555.



7. Eynon N, Hanson ED, Lucia A, et al. Genes for elite power and sprint performance: ACTN3 leads the way. *Sports Med.* 2013; 43: 803-817.
8. Ostrander EA, Huson HJ, Ostrander GK. Genetics of athletic performance. *Annu Rev Genomics Hum Genet.* 2009; 10: 407-429.
9. Peplonska B, Adamczyk JG, Siewierski M, et al. Genetic variants associated with physical and mental characteristics of the elite athletes in the Polish population. *Scand J Med Sci Sports.* 2016; May 3. doi: 10.1111/sms.12687.
10. Cięższyk P, Willard K, Groniek P, et al. Are genes encoding proteoglycans really associated with the risk of anterior cruciate ligament rupture? *Biol Sport.* 2017; 34: 97-103.
11. Reimann F, Cox JJ, Belfer I, et al. Pain perception is altered by a nucleotide polymorphism in SCN9A. *PNAS.* 2010; 107(11): 5148-5153.
12. Pawlak M. Aspects of pain in sport. *Trends Sport Sci.* 2013; 3(20): 123-134.
13. Pawlak M. Praktyczne aspekty sensorycznej i modulującej funkcji nocyceptorów (Practical aspects of the sensory and modulatory function of nociceptors). *Fizjot Pol.* 2008; 2: 115-127.
14. Petho G, Reeh PW. Sensory and signaling mechanisms of bradykinin, eicosanoids, platelet-activating factor, and nitric oxide in peripheral nociceptors. *Physiol Rev.* 2012; 92(4): 1699-1775.
15. Mizumura K, Sugiura T, Katanosaka K, et al. Excitation and sensitization of nociceptors by bradykinin: What do we know? *Exp Brain Res.* 2009; 196(1): 53-65.
16. Rankinen T, Fuku N, Wolfarth B, et al. No evidence of a common DNA variant profile specific to world class endurance athletes. *PLoS One.* 2016; 11(1): e0147330.
17. Iwai N, Ohmichi N, Nakamura Y, Kinoshita M. DD genotype of the angiotensin-converting enzyme gene is a risk factor for left ventricular hypertrophy. *Circulation.* 1994; 90: 2622-2628.
18. Rigat B, Hubert C, Alhenc Gelas F, et al. An insertion/deletion polymorphism in the angiotensin I-converting enzyme gene accounting for half the variance of serum enzyme levels. *J Clin Invest.* 1990; 86: 1343-1346.
19. Williams AG, Rayson MP, Jubbs M, et al. The ACE gene and muscle performance. *Nature.* 2000; 403: 615.
20. Cambien F, Poirier O, Lecerf L, et al. Deletion polymorphism in the gene for angiotensin-converting enzyme is a potent risk factor for myocardial infarction. *Nature.* 1992; 359(6396): 641-644.
21. Norman B, Mahnke-Zizelman DK, Vallis A, Sabina RL. Genetic and other determinants of AMP deaminase activity in healthy adult skeletal muscle. *J Appl Physiol.* 1998; 85: 1273-1278.
22. Fischer H, Esbjörnsson M, Sabina RL, et al. AMP deaminase deficiency is associated with lower sprint cycling performance in healthy subjects. *J Appl Physiol.* 2007; 103: 315-322.
23. Thorstensson A. Muscle strength, fibre types and enzyme activities in man. *Acta Physiol Scand Suppl.* 1976; 443: 1-45.
24. Morisaki T, Sabina RL, Holmes EW. Adenylate deaminase. A multigene family in humans and rats. *J Biol Chem.* 1990; 265(20): 11482.
25. Lowenstein JM. Ammonia production in muscle and other tissues: the purine nucleotide cycle. *Physiol Rev.* 1972; 52(2): 382-414.
26. Rubio JC, Martin MA, Rabadan M, et al. Frequency of the C34T mutation of the AMPD1 gene in world-class endurance athletes: does this mutation impair performance? *J Appl Physiol.* 2005; 98: 2108-2112.
27. Rico-Sanz J, Rankinen T, Joanisse DR, et al. Associations between cardiorespiratory responses to exercise and the C34T AMPD1 gene polymorphism in the HERITAGE Family Study. *Physiol Genomics.* 2003; 14: 161-166.
28. Fedotovskaya ON, Danilova AA, Ahmetov II. Effect of *AMPD1* Gene Polymorphism on Muscle Activity in Humans. *Bull Exp Biol Med.* 2013; 154(10): 485-487.
29. Ginevičienė V, Jakaitienė A, Pranculis A, et al. *AMPD1* rs17602729 is associated with physical performance of sprint and power in elite Lithuanian athletes. *BMC Genet.* 2014; 15: 58.
30. Grenda A, Leońska-Duniec A, Cięższyk P, Zmijewski P. *BDKRB2* GENE -9/+9 polymorphism and swimming performance. *Biol Sport.* 2014; 31(2): 109-113.
31. Regoli D, Barabé J. Pharmacology of bradykinin and related kinins. *Pharmacol Rev.* 1980; 32: 1-46.
32. Saunders CJ, de Milander L, Hew-Butler T, et al. Dipsogenic genes associated with weight changes during Ironman Triathlons. *Hum Mol Genet.* 2006; 15: 2980-2987.
33. Williams AG, Dhamrait SS, Wootton PT, et al. Bradykinin receptor gene variant and human physical performance. *J Appl Physiol.* 2004; 96: 938-942.
34. Fu Y, Katsuya T, Matsuo A, et al. Relationship of bradykinin B2 receptor gene polymorphism with essential hypertension and left ventricular hypertrophy. *Hypertens Res.* 2004; 27: 933-938.
35. Hallberg P, Lind L, Michaëlsson K, et al. B2 bradykinin receptor (B2BKR) polymorphism and change in left ventricular mass in response to antihypertensive treatment: results from the Swedish Irbesartan Left Ventricular Hypertrophy Investigation versus Atenolol (SILVHIA) trial. *J Hypertens.* 2003; 21: 621-624.

36. Braun A, Kammerer S, Maier E, et al. Polymorphisms in the gene for the human B2-bradykinin receptor. New tools in assessing a genetic risk for bradykinin-associated diseases. *Immunopharmacology*. 1996; 33: 32-35.
37. Lung CC, Chan EK, Zuraw BL. Analysis of an exon 1 polymorphism of the B2 bradykinin receptor gene and its transcript in normal subjects and patients with C1 inhibitor deficiency. *J Allergy Clin Immunol*. 1997; 99: 34-46.
38. Holdys J, Gronek P, Kryściak J, Stanisławski D. Genetic variants of uncoupling proteins-2 and -3 in relation to maximal oxygen uptake in different sports *Acta Piochemica Polonica*. 2013; 60(1): 71-75.
39. Fleury C, Sanchis D. The mitochondrial uncoupling protein-2: current status. *Int J Biochem Cell Biol*. 1999; 31(11): 1261-1278.
40. Tu N, Chen H, Winnikes U, et al. Structural organization and mutational analysis of the human uncoupling protein-2 (hUCP2) gene. *Life Sci*. 1999; 64(3): 41-50.
41. Bouchard C, Perusse L, Chagnon YC, et al. Linkage between markers in the vicinity of the uncoupling protein 2 gene and resting metabolic rate in humans. *Hum Molec Genet*. 1997; 6: 1887-1889.
42. Rosmond R, Bouchard C, Björntorp P. Lack of association between the uncoupling protein-2 Ala55Val gene polymorphism and phenotypic features of the metabolic syndrome. *Biochim Biophys Acta*. 2002; 1588(2): 103-105.
43. Warden C. Genetics of uncoupling proteins in humans. *Int J Obes Relat Metab Disord*. 1999; 23(S6): S46-48.
44. Esterbauer H, Schneitler C, Oberkofler H, et al. A common polymorphism in the promoter of UCP2 is associated with decreased risk of obesity in middle-aged humans. *Nature Genet*. 2001; 28: 178-183.
45. Beumann B, Schierning B, Toubro S, et al. The association between the val/ala-55 polymorphism of the uncoupling protein 2 gene and exercise efficiency. *Int J Obes Relat Metab Disord*. 2001; 4: 467-471.
46. Kimm SY, Glynn NW, Aston CE, et al. Racial differences in the relation between uncoupling protein genes and resting energy expenditure. *Am J Clin Nutr*. 2002; 75(4): 714-719.
47. Russell A, Wadley G, Snow R, et al. Slow component of VO<sub>2</sub> kinetics: the effect of training status, fibre type, UCP3 mRNA and citrate synthase activity. *Int J Obes Relat Metab Disord*. 2002; 26(2): 157-164.
48. Schrauwen P, Russell AP, Moonen-Kornips E, et al. Effect of 2 weeks of endurance training on uncoupling protein 3 content in untrained human subjects. *Acta Physiol Scand*. 2005; 183(3): 273-280.
49. Bray M, Hagberg J, Perusse L, Rankinen T, et al. The human gene map for performance and health-related fitness phenotypes: the 2006-2007 update. *Med Sci Sports Exerc*. 2009; 41(1): 34-72. doi: 10.1249/MSS.0b013e3181844179.
50. Culotta E, Koshland Jr DE. NO news is good news. *Science*. 1992; 258(5090): 1862-1865.
51. Moncada S, Higgs A. The L-arginine-nitric oxide pathway. *N Engl J Med*. 1993; 329: 2002-2012.
52. Andrew PJ, Mayer B. Enzymatic function of nitric oxide synthases. *Cardiovasc Res*. 1999; 43: 521-531.
53. Förstermann U, Sessa WC. Nitric oxide synthases: regulation and function. *Eur Heart J*. 2012; 33: 829-837.
54. Lange M, Enkhbaatar P, Nakano Y, Traber DL. Role of nitric oxide in shock: the large animal perspective. *Front Biosci*. 2009; 14: 1979-1989.
55. Harrison DG. Cellular and molecular mechanisms of endothelial cell dysfunction. *J Clin Invest*. 1997; 100: 2153-2157.
56. Hambrecht R, Adams V, Erbs S, et al. Regular physical activity improves endothelial function in patients with coronary artery disease by increasing phosphorylation of endothelial nitric oxide synthase. *Circulation*. 2003; 107: 3152-3158.
57. Kolluru GK, Siamwala JH, Chatterjee S. eNOS phosphorylation in health and disease. *Biochimie*. 2010; 92: 1186-1198.
58. Yang A-L, Tsai S-J, Jyh Jiang M, Chen H. Chronic exercise increases both inducible and endothelial nitric oxide synthase gene expression in endothelial cells of rat aorta. *J Biomed Sci*. 2002; 9(2): 149-155.
59. Gielen S, Sandri M, Erbs S, Adams V. Exercise-induced modulation of endothelial nitric oxide production. *Curr Pharm Biotechnol*. 2011; 12: 1375-1384.
60. Green DJ, Spence A, Halliwill JR, et al. Exercise and vascular adaptation in humans. *Exp Physiol*. 2011; 96: 57-70.
61. Kojda G, Cheng YC, Burchfield J, Harrison DG. Dysfunctional regulation of endothelial nitric oxide synthase (eNOS) expression in response to exercise in mice lacking one eNOS gene. *Circulation*. 2001; 103: 2839-2844.
62. Pospieszna B, Karolkiewicz J, Tarnas J, et al. Influence of 12-week Nordic Walking training on biomarkers of endothelial function in healthy postmenopausal women. *J Sports Med Phys Fit*. 2016; Sep 22.
63. Marsden PA, Heng HH, Scherer SW, et al. Structure and chromosomal localization of the human constitutive endothelial nitric oxide synthase gene. *J Biol Chem*. 1993; 268(23): 17478-17488.
64. Vecoli C. Endothelial nitric oxide synthase gene polymorphisms in cardiovascular disease. *Vitam Horm*. 2014; 96: 387-406.